FILE 'HOME' ENTERED AT 15:51:05 ON 22 APR 2008

=> index bioscience

FILE 'DRUGMONOG' ACCESS NOT AUTHORIZED

COST IN U.S. DOLLARS

SINCE FILE ENTRY SESSION 0.21 0.21

TOTAL

FULL ESTIMATED COST

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, ANTE, AQUALINE, AQUASCI, BIOENG, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CAPLUS, CEABA-VTB, CIN, CONFSCI, CROPB, CROPU, DDFB, DDFU, DGENE, DISSABS, DRUGB, DRUGMONOG2, DRUGU, EMBAL, EMBASE, ...' ENTERED AT 15:51:17 ON 22 APR 2008

69 FILES IN THE FILE LIST IN STNINDEX

Enter SET DETAIL ON to see search term postings or to view search error messages that display as 0* with SET DETAIL OFF.

- => silk and chloroform and hydrogel
 - FILE CAPLUS 1
 - 1 FILE IFIPAT
 - FILE USPATFULL 419
 - 75 FILE USPAT2
 - 65 FILES SEARCHED...
 - 1 FILE WPIDS
 - FILE WPINDEX
 - 6 FILES HAVE ONE OR MORE ANSWERS, 69 FILES SEARCHED IN STNINDEX
- QUE SILK AND CHLOROFORM AND HYDROGEL T.1
- => silk and hexane and hydrogel
 - 1 FILE CAPLUS
 - FILE IFIPAT 3
 - 395 FILE USPATFULL
 - 103 FILE USPAT2
 - FILE WPIDS 2
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 - 6 FILES HAVE ONE OR MORE ANSWERS, 69 FILES SEARCHED IN STNINDEX
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- => silk and iso-amyl and hydrogel
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 - FILE USPAT2 10
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- QUE SILK AND ISO-AMYL AND HYDROGEL
- => silk or collagens or keratins or actins or chorions or seroins
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 - FILE ADISINSIGHT 6
 - FILE ADISNEWS 4
 - 2724 FILE AGRICOLA

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FILE TOXCENTER

FILE USPATFULL

FILE USPATOLD

FILE USGENE

FILE USPAT2 FILE VETB

FILE VETU FILE WATER

17916 FILE WPIDS

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29444

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84

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325 FILE WPIFV
17916 FILE WPINDEX
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66 FILES HAVE ONE OR MORE ANSWERS, 69 FILES SEARCHED IN STNINDEX

L4 OUE SILK OR COLLAGENS OR KERATINS OR ACTINS OR CHORIONS OR SEROINS

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            FILE USPAT2
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  6 FILES HAVE ONE OR MORE ANSWERS, 69 FILES SEARCHED IN STNINDEX
L8 QUE L6 AND HYDROGEL
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          10 USPAT2
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           1 CAPLUS
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           1 IFIPAT
F5
           1 WPIDS
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           1 WPINDEX
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COST IN U.S. DOLLARS
FULL ESTIMATED COST
                                                    9.10 9.31
FILE 'CAPLUS' ENTERED AT 15:59:35 ON 22 APR 2008
USE IS SUBJECT TO THE TERMS OF YOUR STN CUSTOMER AGREEMENT.
PLEASE SEE "HELP USAGETERMS" FOR DETAILS.
COPYRIGHT (C) 2008 AMERICAN CHEMICAL SOCIETY (ACS)
FILE 'IFIPAT' ENTERED AT 15:59:35 ON 22 APR 2008
COPYRIGHT (C) 2008 IFI CLAIMS(R) Patent Services (IFI)
FILE 'WPIDS' ENTERED AT 15:59:35 ON 22 APR 2008
COPYRIGHT (C) 2008 THE THOMSON CORPORATION
=> 16 and hydrogel
L9
   3 L6 AND HYDROGEL
=> d ab bib 1-3
L9 ANSWER 1 OF 3 CAPLUS COPYRIGHT 2008 ACS on STN
AB One aspect of the present invention relates to a method of preparing a
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fibrous protein smectic hydrogel by way of a solvent templating process, comprising the steps of pouring an aqueous fibrous protein solution into

a container comprising a solvent that is not miscible with water; sealing the container and allowing it to age at about room temperature; and collecting the resulting fibrous protein smectic hydrogel and allowing it to dry. Another aspect of the present invention relates to a method of obtaining predominantly one enantiomer from a racemic mixture, comprising the steps of pouring an aqueous fibrous protein solution into a container comprising a solvent that is not miscible with water; sealing the container and allowing it to age at about room temperature; allowing the enantiomers of racemic mixture to diffuse selectively into the smectic hydrogel in solution; removing the smectic hydrogel from the solution; rinsing predominantly one enantiomer from the surface of the smectic hydrogel; and extracting predominantly one enantiomer from the interior of the smectic hydrogel. The present invention also relates to a smectic hydrogel prepared according to an aforementioned method.

AN 2004:412959 CAPLUS

DN 140:420370

TI Templated native silk smectic gels

IN Valluzzi, Regina; Jin, Hyoung-Joon; Park, Jaehyung

PA Trustees of Tufts College, USA

SO PCT Int. Appl., 66 pp.

CODEN: PIXXD2

DT Patent

LA English

FAN.CNT 1

r An.	PATENT NO.									APPLICATION NO.									
PI		2004041845 2004041845				A2				WO 2003-US34684					20031031				
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PRAI	JP US IN US	R: AT, BE, CH IE, SI, LT 1732014 2006504852 20060205927 2005CN01071 2002-423046P 2003-US34684			LT,	LV, A T A1 A	FI,	RO, 2006 2006 2006 2007 2002	MK, 0208 0209 0914 0824 1101	CY,	AL, CN 2 JP 2 US 2	TR, 1003- 1004- 1005-	BG, 8010 5503 5336	CZ, 7979 38 11	EE,	HU, 2: 2:	SK 0031 0031 0050	031 031 429	

L9 ANSWER 2 OF 3 IFIPAT COPYRIGHT 2008 IFI on STN

AB One aspect of the present invention relates to a method of preparing a fibrous protein smectic hydrogel by way of a solvent templating process, comprising the steps of pouring an aqueous fibrous protein solution into a container comprising a solvent that is not miscible with water; sealing the container and allowing it to age at about room temperature; and collecting the resulting fibrous protein smectic

hydrogel and allowing it to dry. Another aspect of the present invention relates to a method of obtaining predominantly one enantiomer from a racemic mixture, comprising the steps of pouring an aqueous fibrous protein solution into a container comprising a solvent that is not miscible with water; sealing the container and allowing it to age at about room temperature; allowing the enantiomers of racemic mixture to diffuse selectively into the smectic hydrogel in solution; removing the smectic hydrogel from the solution; rinsing predominantly one enantiomer from the surface of the smectic hydrogel; and extracting predominantly one enantiomer from the interior of the smectic hydrogel. The present invention also relates to a smectic hydrogel prepared according to an aforementioned method. 11256867 IFIPAT; IFIUDB; IFICDB TEMPLATED NATIVE SILK SMECTIC GELS

ΑN

ΤI

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PΙ US 2006205927 A1 20060914 ΑI US 2003-533611 20031031 WO 2003-US34684 20031031

20060511 PCT 371 date 20060511 PCT 102(e) date

PRAI US 2002-423046P 20021101 (Provisional)

FΙ US 2006205927 20060914

DT Utility; Patent Application - First Publication

FS CHEMICAL APPLICATION

EDEntered STN: 15 Sep 2006 Last Updated on STN: 15 Sep 2006

The invention was made with support provided by NASA (grant NAG81699) and NSF (grant BES 9727401); therefore, the government has certain rights in the invention.

CLMN

GT 33 Figure(s).

- FIG. 1 depicts the surface (right) and fracture surface of chloroform templated silk. The wavy texture is everywhere on the solvent templated side of the materials surface.
- FIG. 2 depicts the chloroform templated film. Waves reorienting and becoming terraces can be seen, behavior which is not expected for simple wrinkles due to contraction.
- FIG. 3 depicts a regular pattern of nubby small structures comprising the waves.
- FIG. 4 depicts an amyl alcohol film showing a surface that looks like a "nonwoven woven" fabric.
- FIG. 5 depicts a surface texture seen at an angle showing a thin layer very different from the chloroform films.
- FIG. 6 depicts amyl alcohol templated samples soaked in bipyridyl trisRuII chloride hexahydrate giving a high magnification image and a 40 nm layered feature.
- FIG. 7 depicts films after soaking in a dysprosium chloride solution for added contrast. The wavy layered structure of the chloroform templated film is apparent here.
- FIG. 8 depicts a film's texture that is even and regular.
- FIG. 9 depicts self-fabricated textured "tapes" from a peptide with sequence (Glu)5(Ser-Gly-Ala-Gly-Val-Gly-Arg-Gly-Asp-GlySer-GlyVal-Gly-Leu-

- Gly-Ser-Gly-Asn-Gly)2(Glu)5. 1. Optical micrograph shows a 10-15 micron texture which persists through the material thickness. The material is optically transparent. 2. Polarizing optical microscopy reveals patterned birefringence, indicating that the topographic texture is due to a changing material orientation. 3. SEM image shows the topographic structure of the tape. The difference in periodicity observed in SEM and optical microscopy is due to the fact that top surface and bottom surface ridges are both observed in the optical image (resulting in an apparently shorter period).
- FIG. 10 depicts self-fabricated tapes of (Glu)5(Ser-Gly-Ala-GlyVal-Gly-Arg-Gly-Asp-Gly-Ser-Gly-Val-Gly-Leu-Gly-Ser -Gly-Asn-Gly) 2(Glu)5 have "patterns within patterns" or a long-range ordered structure consisting of hierarchical nanoscale to microscale patterns; 1: the self-limited width and thickness of the fibers (120 microns, 50 microns respectively) form the largest length scale in the hierarchy; a 40 micron periodic texture is observed running along the tape; 2: within the ridges of the 40 micron texture a 3 micron subtexture is observed; 3: a submicron texture of inclined sheets or layers can be observed (<40 nm, but exact size is below the resolution of the scanning electron microscope); TEM studies indicate a layer spacing of 5 nm.
- FIG. 11 depicts an IR spectra of self-fabricated tapes of (Glu) 5(Ser-Gly-Ala-Gly-Val-Gly-Arg-Gly-Asp-Gly-Ser-GlyVal-Gly-LeuGly-Ser-Gly-Asn-Gly)2(Glu)5. Typically IR spectra for molecules are seen as very small differences in IR transmission relative to a large backround, which must be subtracted out Raw data (no background subtraction) is shown for transmission FTIR spectra through different regions (orientations) of the tape structure. Two orientations show very typical protein absorbance spectra over a high background. However in some orientations the IR radiation does not reach the detector.
- FIG. 12 depicts an IR spectrum modified by tape with scale expanded to show spectral features. Instead of an absorption or transmission spectrum, a pattern of 2 overlaid sinusoids (one has a 50/cm period, the other a 25/cm period. The effect for this material appears strongest in the 1750-3500 cm-1, or 5.7-2. 9 micron range.
- FIG. 13 depicts twisted polycrystals obtained by salt precipitation of an oligopeptide with Na-EDTA.
- FIG. 14 depicts ordered "corkscrew" polycrystalline oligopeptide salt precipitate as a hierarchy of twisted ordered structures.
- FIG. 15 depicts reflection and transmission FTIR spectra for ordered polycystals. TOP: reflection infrared spectrum, Raw data. A glassy disordered material of the oligopeptide is more reflective than the background. An ordered periodic nanolayered material from the same peptide is shown, and clearly reflects far less of the infrared radiation. BOTTOM: transmission spectra for background, unordered peptide material and a chemically identical nanolayered ordered material of the peptide. Spectrum is greatly attenuated for the ordered material.
- FIG. 16 depicts ordered textured surfaces and interiors from templated gels (a) chloroform templated gels have a wavy surface texture covering the surfaces which were in contact with water; (b) a fracture surface from the chloroform templated gel reveals a "skin" of the wavy pattern, which forms channels down into the interior, the interior has a different structure, which appears to be made of wavy plates; (c) templated surface of amyl alcohol templated material (in contact with water); (d) higher magnification image of the edge of the region in c, showing a "skin" core structure and a patterned texture throughout the material; (e,f) amyl alcohol dried film after swelling in an aqueous solution of ruthenium compound and extraction of ruthenium compound by swelling in water; (e) wavy lines indicate reorientation of ordered structures within the material; (f) at high magnification (20, 000x) lines 38 nm in width are observed.
- FIG. 17 depicts amyl alcohol templated gel after soaking in Aqueous

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Tris(2,2'-bipyridyl) dichloro ruthenium(II) hexahydrate ("Rubipy")
      solution for 1 day. Much of the Rubipy has migrated from the solution
      into the silk gel. Initial migration is rapid and chirally
      selective (occurs over roughly 1 hour). Additional migration occurs
      slowly after this for roughly 1 day and is less chirally selective.
     Chloroform templated gels do not exhibit complex diffusion behavior and
      are chirally selective throughout the swelling process.
     FIG. 18 depicts a cross section of amyl alcohol templated gel after
      swelling in Rubipy for 1 hour. The Rubipy penetrated rapidly into the
      outer "skin" layers of the gel (bright orange), and more slowly into the
      interior (yellowish region).
     FIG. 19 depicts an X-ray diffraction pattern from chloroform templated
     gel. Dark arcs along the diffraction rings (arrow) indicate orientation.
     FIG. 20 depicts the non-globular nature of fibrous proteins.
     FIG. 21 depicts the long range order of liquid crystals.
     FIG. 22 depicts "frustration" in nanolayered crystals.
     FIG. 23 depicts nanocomposites.
     FIG. 24 depicts banded structures from native silk.
     FIG. 25 depicts banded structures from engineered protein designed
     peptide.
     FIG. 26 depicts how hairpin structures allow silk liquid
     crystallinity.
     FIG. 27 depicts spider silk modification.
     FIG. 28 depicts amphiphilic spider silk motif.
     FIG. 29 depicts silkworm silk peptide models.
     FIG. 30 depicts film morphology and helix anchoring.
     FIG. 31 depicts the templating-against-solvent technique.
     FIG. 32 depicts patterned peptide films.
     FIG. 33 depicts silk templated gels-surface "skin".
    ANSWER 3 OF 3 WPIDS COPYRIGHT 2008
                                               THE THOMSON CORP on STN
    WO 2004041845 A2
                      UPAB: 20060121
     NOVELTY - Preparing a fibrous protein smectic hydrogel
     comprises:
          (1) pouring an aqueous fibrous protein solution into a container
     comprising a solvent that is not miscible with water;
          (2) sealing the container and allowing it to age at about room
     temperature; and
          (3) collecting the resulting fibrous protein smectic hydrogel
     and allowing it to dry.
            DETAILED DESCRIPTION - An INDEPENDENT CLAIM is included for a
     method of obtaining predominantly one enantiomer from a racemic mixture.
            USE - The method is useful in preparing a fibrous protein smectic
     hydrogel (claimed).
     2004-411501 [38]
                        WPIDS
DNC C2004-154468 [38]
     Preparing a fibrous protein smectic hydrogel by pouring an
     aqueous fibrous protein solution into a container, sealing the container
     and allowing it to age at room temperature and collecting and allowing to
     dry the resulting hydrogel
     JIN H; JIN H J; PARK J; VALLUZZI R
     (TUFT-C) TUFTS COLLEGE; (TUFT-C) UNIV TUFTS
    105
    WO 2004041845
                    A2 20040521 (200438)* EN
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     AU 2003294240
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     US 20060205927 A1 20060914 (200661)
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ADT WO 2004041845 A2 WO 2003-US34684 20031031; US 20060205927 A1 Provisional US 2002-423046P 20021101; AU 2003294240 A1 AU 2003-294240 20031031; CN 1732014 A CN 2003-80107979 20031031; EP 1565203 A2 EP 2003-789721 20031031; EP 1565203 A2 WO 2003-US34684 20031031; JP 2006504852 W WO 2003-US34684 20031031; US 20060205927 A1 WO 2003-US34684 20031031; JP 2006504852 W JP 2004-550338 20031031; US 20060205927 A1 US 2006-533611 20060511; IN 2005CN01071 P4 WO 2003-US34684 20031031; IN 2005CN01071 P4 IN 2005-CN1071 20050531

FDT AU 2003294240 A1 Based on WO 2004041845 A; EP 1565203 A2 Based on WO 2004041845 A; JP 2006504852 W Based on WO 2004041845 A

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